

International Journal of Current Microbiology and Applied Sciences ISSN: 2319-7706 Volume 7 Number 05 (2018) Journal homepage: <u>http://www.ijcmas.com</u>



Original Research Article

https://doi.org/10.20546/ijcmas.2018.705.316

Antibacterial Efficacy and Cluster Analysis of Genotypic Extracts of Coriander Leaves and Seeds against Human Pathogenic Bacterial Strains

Ankita Verma*, Deepali Agarwal, Jyotsna Dhanik, Neelam Arya and Viveka Nand

Department of Chemistry, College of Basic Sciences and Humanities, G.B. Pant University of Agriculture and Technology, Pantnagar – 263145, Uttarakhand, India

*Corresponding author

The present study was undertaken to evaluate the in vitro antibacterial activity of

seventeen genotypes of Coriander leaves and seeds collected from Tarai and Kumaun regions of Uttarakhand state of India. The methanolic extracts of leaves and seeds

genotypes were screened against four human pathogenic bacterial strains; Escherichia coli,

Bacillus subtilis, Pseudomonas aeruginosa, Salmonella typhi at four different

concentrations. All the methanolic extracts of leaves and seeds had inhibitory effect against tested bacteria *E. coli*, *S. typhi* and *B. subtilisi*, while no antibacterial activity was

observed against P. aeruginosa for both leaves and seeds extracts. The study also evaluates

the genotypic variation and similarities that exist between the Coriander leaves and seeds

on the basis of its antibacterial potency through a statistical approach i.e. cluster analysis,

regarding genetic makeup of Coriander cultivars so that future investigation may be

carried out to develop a new variety for further applications in pharmacology.

ABSTRACT

Keywords

Coriander, genotypes, Escherichia coli, Bacillus subtilis, Pseudomonas aeruginosa, Salmonella typhi, cluster analysis

Article Info

Accepted: 20 April 2018 Available Online: 10 May 2018

Introduction

Spices are products from plant leaves, seeds, fruits, flowers, roots or bark, that are added to food commodities to enhance flavour, colour or to minimize the rate of rancidity and as preservatives that suppress microbial activities (Teye et al.. 2013). The beneficial characteristics of natural plant products could increase food safety and shelf life of processed food products (Dhanik et al., 2017). The benefits of these herbs and spices are not hidden and it is only in recent years that modern science has started paying attention to the properties of spices (Chaudhry and Tariq, 2006). These plants then emerged as

compounds with potentially significant therapeutic application against human pathogens, including bacteria, fungi or virus (Horvath *et al.*, 2002; Jantan *et al.*, 2003; Khan *et al.*, 2003; Perez, 2003).

These traditional medicinal plants contain a wide range of substances that can be used to treat chronic as well as infectious diseases (Reynolds, 1996). The antibacterial activity of various spices and medicinal plants has been known for years. Plant oils and extracts have been used for a wide variety of purposes for many thousands of years (Hashim *et al.*, 2010). The use of these plants containing natural compounds could open up the

possibility of using them as novel antimicrobials (Dhanik *et al.*, 2017).

One of the widely used spices having tremendous medicinal properties is Coriander. Coriander, also called as "cilantro", is an annual herbaceous plant of family Apiaceae originating from Middle Eastern and Mediterranean regions cultivated for its aromatic and medicinal use (Mildner et al., 2009). It is considered both as an herb and a spice since both of its leaves and seeds are used as a seasoning condiment. Researches on potential botanical extracts which are safe and environmentally benign are urgently needed. The medicinal properties of Coriander may be attributed due to the presence of various phytoconstituents present in its leaves and seeds. In our previous work (Verma et al., 2018), we have investigated that the type of phytoconstituents of leaves and seeds of different genotypes are not similar. The use of this magical plant is not only limited to household health benefits rather it is also used in agricultural field for control of insect and pests due to the presence of plant secondary constituents, that plays a prominent role with different mode of action against insects. Genotypic extracts of Coriander leaves and seeds were also tested as antifeedants against insect S. litura (Verma et al., 2015). Coriander is also used in detox diet as it helps to remove toxic mineral residue such as mercury and lead, and excrete them in the urine or faeces (Kansal et al., 2011). Coriander is considered an important herb due to its extensive use as medicine for curing capabilities against many diseases due to presence of active ingredients and components in its leaves and fruit (Kubo et al., 2004).

The mountainous state of Uttarakhand, India, is endowed with a rich variety of medicinal plants, many of which are in great demand in the domestic and export markets (Government of India, 2000). Because of a variety of agroclimatic niches, the state has tremendous potential to emerge as a regular supplier of medicinal and aromatic plants (Samant *et al.*, 1998). Uttarakhand State has a rich heritage of traditional medicine system that acquires a dominant role in pharmacy and agronomy production. Pharmacology, phytochemistry and biological activities of some medicinal plant of Uttarakhand have been reported earlier by Dhanik *et al.*, (2017).

The aim of our study is to investigate the antibacterial activity of different genotypic extracts of both seeds and leaves of Coriander cultivars against four human pathogenic bacterial strains. In addition, cluster analysis was applied to achieve the knowledge about the genetic makeup of Coriander cultivars and to develop a new variety of Coriander for future pharmacological applications.

Materials and Methods

Plant material

Seventeen different genotypes of Coriander were developed and collected from Vegetable Research Centre (V.R.C) of G.B.P.U.A & T, Pantnagar and from Kumaun hills. Uttarakhand, India. Out of the total seventeen genotypes collected for experimental analysis, fifteen genotypic varieties were developed in Pantnagar Tarai area viz. Pant haritima, PD-21, PD-51, UD-643, UD-684, UD-699, UD-704, UD-711, UD-716, UD-720, UD-721, UD-722, UD-725, UD-727, UD-728 and two were collected from Kumaun Region viz. PD 52, Pithoragarh Region and PD 53, Berinag Region of Uttarakhand State.

Preparation of plant extracts

The collected samples of Coriander leaves were washed in a running tap to remove soil and dust particles and then shade dried in the laboratory for seven days. The dried samples of Coriander leaves and seeds were pulverized into fine powder with a mechanical grinder. The material was extracted by successive soaking for 72 hours in methanol and stored in refrigerator at 4°C in a dry, clean amber coloured bottle for further analysis. Working standards of desired concentration 250, 500, 750 and 1000 μ g/mL of methanolic extracts were then prepared each time from the stock to conduct the experiment.

Sources of test organisms

The antibacterial screening of methanolic leaves and seeds extracts of Coriander was evaluated genotypes against four pathogenic bacterial strains; Escherichia coli, Salmonella typhi, Bacillus subtilis. Pseudomonas aeruginosa. The bacterial strains used for antibacterial study was isolated from different meat samples in Department of Veterinary and Public Health, Pantnagar (India). Antibacterial screening of the extracts against these bacteria was done by Disc-diffusion method as reported by Singh et al., (2005) with slight modification and was measured by zone of inhibition.

Preparation of bacterial inoculums

For the preparation of bacterial inoculums, Luria Bertani broth (Hi-media) for Escherichia coli, buffered peptone water (Himedia) for Salmonella typhi, nutrient broth for Bacillus subtilis and Pseudomonas aeruginosa were weighed and poured in distilled water as per manufacturer's instructions. The test tubes containing culture media was sterilized in an autoclave at 120°C and 15-20 lbs for half an hour. Bacterial colonies were inoculated in test tubes in above prepared broths. The test tubes containing bacterial colonies were incubated for 24 hr in incubator. Next day cultures showed a marked turbidity in the tubes and were used to conduct further experiment.

Preparation of agar plates

Difco nutrient agar (1.5%) was used for the preparation of plate media. The media was prepared in distilled water, autoclaved and gently cooled. Thereafter, the prepared media was poured in petri plates (dia 9 cm) in laminar flow and kept undisturbed as such till it got solidified. After solidification, these petri plates were incubated at 37°C overnight for sterile testing.

Screening of antibacterial activity of methanolic extracts of coriander genotypes

The standard disc diffusion method was used for the antibacterial analysis (Bauer *et al.*, 1966; NCCLS, 2000). Bacterial inoculums 100 μ L was added to the agar plates and uniformly spread over the surface using spreader (Hi-media).

Placement of the plates

Sterilized disc of 5 mm diameter soaked in different methanolic concentrations of extracts (250, 500, 750 and 1000 µg/mL) were placed on the inoculated plate. These plates were incubated at 37°C overnight to observe the zone of inhibitions around the disc, which were compared with the zone of inhibitions formed by the standard antibiotic. Gentamicine. The sterile paper discs impregnated with methanol served as negative control. These inoculated plates were firmly closed with the maximum possible aseptic precautions.

Recording of observations

After incubation, relative susceptibility of each organism was determined by a clear zone of inhibition of growth around the disc impregnated with the extracts as well as the antibiotic. Zone of inhibition (mm) was measured with the help of scale.

Statistical analysis

Cluster analysis was performed using the software The Unscrambler X 10.5 applying Ward's method with Square Euclidean distance.

Results and Discussion

The antibacterial screening of various concentrations of methanolic extracts of Coriander leaves and seeds genotypes against four human pathogenic bacterial strains; *Escherichia coli, Salmonella typhi, Bacillus subtilis and Pseudomonas aeruginosa* was performed by Disc-diffusion method and antibacterial activity was measured as zone of inhibition.

All the methanolic extracts of leaves and seeds had inhibitory effect against tested bacteria; *E. coli*, *S. typhi and B. subtilis* while no antibacterial activity was observed with *P. aeruginosa* for both leaves and seeds extracts.

Methanolic extract of PD-51 leaves and Pant haritima seeds exhibited maximum zone of inhibition for all the bacterial strains. Leaves extracts of UD-684, UD-704, UD-711, UD-716, UD-721 and PD-52 were found to be ineffective against *B. subtilis.* Similarly, seeds extracts of UD-720 and UD-727 showed no activity against *S. typhi.*

The zone of inhibition for Coriander leaves and seeds genotypes as recorded for different bacteria are presented in table 1 and 2, figure 1 and 2. The inhibitory effect of Coriander genotypes was comparatively less than that of standard antibiotic, Gentamicin. Ali *et al.*, (2008) reported that the antimicrobial efficiency of Coriander leaf extracts is due to the presence of flavonoids and terpenoids. The essential oil and various extracts from Coriander have been shown to possess antibacterial, antidiabetic, anticancerous,

antimutagenic, antioxidant and free radical scavenging activities (Sreelatha et al., 2009; Zoubiri and Baaliouamer, 2010). It is also reported that Coriander essential oil has powerful antibacterial activity against gram positive (Staphylococcus aureus and Bacillus spp.) and gram negative (Escherichia coli, Salmonella typhi, Klebsiella pneumonia and Proteus mirabilis) bacteria (Cantore et al., 2004; Matasyoh et al., 2009). From the present study, we conclude that methanolic extracts of Coriander leaves and seeds have pronounced antibacterial potency and respond differently to different bacteria with variable zone of inhibition. Elgayyae et al., (2001) reported that the antibacterial effect of Coriander appeared to be highly variable depending on plants conditions and species.

Cluster analysis

Cluster analysis applied on the entire data including all the concentrations of methanolic extracts of leaves and seeds of Coriander genotypes, gave respective dendrograms presented in figure 3 and 4, where prefix L and S denotes leaves and seeds genotypes respectively. Based on the antibacterial efficacy of Coriander genotypes as screened against bacterial strains various genotypes of Coriander leaves and seeds were grouped into five clusters each as:

Clustering in leaves genotypes

Group 1: L-UD-699, L-UD-725, L-PD-53

Group 2: L-UD-722, L-UD-728, L-UD-720, L-PD-51

Group 3: L-UD-643, L-Pant haritima

Group 4: L-UD-704, L-UD 684, L-PD-21

Group 5: L-UD-721, L-UD-716, L-UD-711, L-UD-727, L-PD-52

Int.J.Curr.Microbiol.App.Sci (2018) 7(5): 2727-2736

Leaves Genotypes	Zone of inhibition (mm)															
	Ì	Escherichia coli			Salmonella typhi					Bacillu	s subtil	is	Pseudomonas aeruginosa			
	Conc.(ppm)			Conc. (ppm)					Conc	(ppm))	Conc. (ppm)				
	250	500	750	1000	250	500	750	1000	250	500	750	1000	250	500	750	1000
Pant haritima	-	12	12	14	10	12	-	-	12	12	-	12	-	-	-	-
PD-21	-	-	-	16	-	10	12	12	-	-	-	12	-	-	-	-
PD-51	10	16	18	20	-	-	-	14	-	12	12	14	-	-	-	-
PD-52	10	10	12	12	-	-	10	12	-	-	-	-	-	-	-	-
PD-53	10	10	10	-	-	10	10	10	12	-	10	14	-	-	-	-
UD-643	-	16	16	-	12	10	-	-	10	10	12	16	-	-	-	-
UD-684	-	-	-	-	-	10	12	12	-	-	-	-	-	-	-	-
UD-699	10	10	10	14	-	10	14	-	12	12	14	-	-	-	-	-
UD-704	-	-	-	12	-	12	10	-	-	-	-	-	-	-	-	-
UD-711	14	12	14	16	10	-	-	10	-	-	-	-	-	-	-	-
UD-716	10	10	10	12	16	12	12	10	-	-	-	-	-	-	-	-
UD-720	-	14	14	20	-	8	10	12	-	10	10	12	-	-	-	-
UD-721	12	10	10	-	-	10	10	16	-	-	-	-	-	-	-	-
UD-722	-	10	10	10	-	12	12	12	-	-	14	14	-	-	-	-
UD-725	10	12	12	14	-	10	12	12	10	-	-	12	-	-	-	-
UD-727	-	10	12	16	-	-	-	14	-	-	-	-	-	-	-	-
UD-728	-	12	12	12	-	8	10	12	12	12	14	16	-	-	-	-
Gentamicine	24				24					2	24		20			
Control (Methanol)	-				-						-		-			

Table.1 Effect of Coriander leaves genotypes on human pathogenic bacteria by disc diffusion method

(-) sign indicates no inhibition

Table.2 Effect of coriander seeds genotypes on human pathogenic bacteria by disc diffusion method

Seeds Genotypes	Zone of inhibition (mm)															
		Escher	ichia co	oli	Salmonella typhi				Bacillus subtilis				Pseudomonas aeruginosa			
		Conc	.(ppm)		Conc. (ppm)				Conc. (ppm)				Conc. (ppm)			
	250	500	750	1000	250	500	750	1000	250	500	750	1000	250	500	750	1000
Pant haritima	-	-		16	12	12	16	16	-	10	12	14	-	-	-	-
PD-21	10	12	12	14	-	12	12	14	-	10	10	12	-	-	-	-
PD-51	-	10	12	14	10	10	10	12	10	12	12	14	-	-	-	-
PD-52	10	12	14	16	-	-	-	10	12	12	12	14	-	-	-	-
PD-53	10	14	14	16	-	10	10	12	-	12	14	14	-	-	-	-
UD-643	-	12	14	16	10	12	12	14	-	-	-	10	-	-	-	-
UD-684	-	-	-	-	-	12	14	14	-	-	-	12	-	-	-	-
UD-699	10	12	12	14	-	10	12	14	10	10	12	12	-	-	-	-
UD-704	-	-	-	12	-	10	10	12	-	10	10	12	-	-	-	-
UD-711	-	14	14	16	12	12	14	16	12	12	14	16	-	-	-	-
UD-716	-	12	12	14	10	12	14	14	10	10	12	12	-	-	-	-
UD-720	-	14	14	16	-	-	-	-	-	10	10	12	-	-	-	-
UD-721	10	12	12	14	-	12	16	16	-	12	14	14	-	-	-	-
UD-722	10	12	12	12	-	10	12	12	12	12	12	14	-	-	-	-
UD-725	-	10	12	14	-	10	10	10	10	10	12	14	-	-	-	
UD-727	-	12	14	14	-	-	-	-	-	-	-	12	-	-	-	-
UD-728	-	14	14	16	-	10	12	12	-	-	-	14	-	-	-	-
Gentamicine			24			1	24				24		20			
Control (Methanol)			-				-				-		-			

(-) sign indicates no inhibition

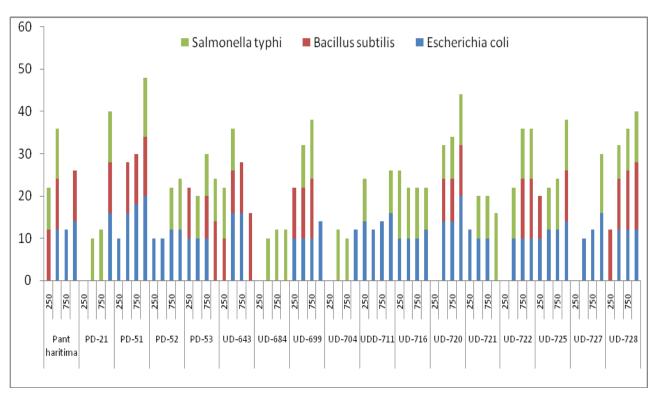
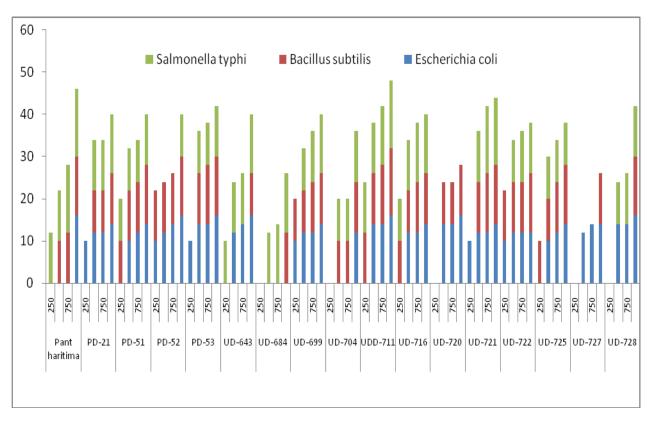


Fig.1 Zone of inhibition in different bacterial strains by coriander leaves genotypes

Fig.2 Zone of inhibition in different bacterial strains by coriander seeds genotypes



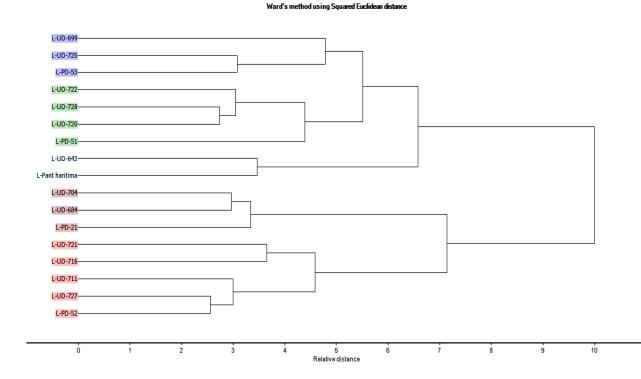
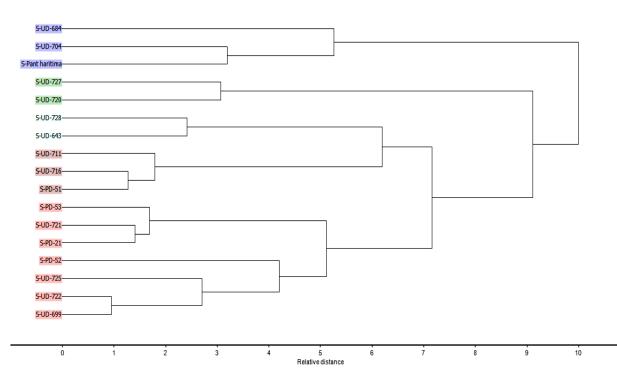


Fig.3 Dendrogram of various genotypes of coriander leaves grouped into five clusters

Fig.4 Dendrogram of various genotypes of coriander seeds grouped into five clusters



Ward's method using Squared Euclidean distance

Clustering in seeds genotypes

Group 1: S-UD-684, S-UD-704, S-Pant haritima

Group 2: S-UD-727, S-UD-720

Group 3: S-UD-728, S-UD-643

Group 4: S-UD-711, S-UD-716, S-PD-51

Group 5: S-PD-53, S-UD-721, S-PD-21, S-PD-52, S-UD-725, S-UD-722, S-UD-699

This scientific and statistical information provides an important platform for the development of effective natural medicines by developing clusters of Coriander genotypes which can serve as a potential natural antibacterial plant extract for bacteria's which are threat to human health.

To the best of our knowledge, this is the first report on antibacterial efficacy of Coriander leaves and seeds genotypes developed and collected from Tarai and Kumaun region of Uttarakhand state and present study results suggest that this new investigation could be first step to develop a new variety of Coriander leaves and seeds for future applications pharmacology. in Cluster analysis provides five clusters of respective genotypes of Coriander leaves and seeds depending on their potency of antibacterial activity irrespective of concentration. This study provides evidence for further statistical approach to identify the active components responsible for the plant biological activity.

Acknowledgement

Authors are sincerely thankful to Department of Science and Technology, New Delhi for the award of DST-INSPIRE Fellowship. Vegetable Research Centre and Department of Veterinary and Public Health (G.B.P.U.A&T, Pantnagar, India) are also thankfully acknowledged.

References

- Ali, S.S., Kasoju, N. and Luthra, A. (2008).
 Indian medicinal herbs as sources of antioxidants. Food Research International. 41: 1-15.
- Bauer, A.W., Kirby, W.M., Sherris, J.C. and Turck, M. (1966). Antibiotic susceptibility testing by a standardized single disc method. American Journal of Clinical Pathology. 45: 493-496.
- Cantore, P.L., Iacobellis, N.S., Marco, A.D., Capasso, F. and Senatore, F. (2004). Antibacterial activity of *Coriandrum* sativum L. and *Foeniculum vulgare Miller Var. vulgare* (Miller) essential oils. Journal of Agricultural and Food Chemistry. 52: 7862-7866.
- Chaudhry, N.M.A. and Tariq, P. (2006). Bactericidal activity of black peeper, bay leaf, aniseed and Coriander against oral isolates. Pakistan Journal of Pharmaceutical Sciences. 19: 214-218.
- Dhanik, J., Verma, A., Arya, N., Verma, A. and VivekaNand. (2017). A brief review on some medicinal plants of Uttarakhand. Journal of Pharmacognosy and Phytochemistry. 6 (6): 2497-2506.
- Dhanik, J., Verma, A., Arya, N., and VivekaNand. (2017). Chemical Profiling and Antioxidant Activity of Essential Oil of *Zingiber officinale* Roscoe from Two Different Altitudes of Uttarakhand. Journal of Essential Oil Bearing Plants. 20 (6): 1547-1556.
- Dhanik, J., Verma, A., Arya, N., Prakash, O. and VivekaNand. (2017). Chemical Profiling and Antibacterial Efficacy of Different Ginger Accessions from Uttarakhand, India. Asian Journal of Chemistry. 29 (10): 2123-2128.
- Elgayyar, M., Draughon, F.A., Golden, D.A. and Mount, J.R. (2001). Antimicrobial

of essential oils from plants against selected pathogenic and saprophytic microorganisms. Journal of Food Protection. 64(7): 1019-1024.

- Government of India (2000). Report of the task force on conservation and sustainable use of medicinal plants (New Delhi: Planning Commission).
- Hashim, H., Kamali, E.L. and Mohammed, Y. (2010). Antibacterial Activity and Phytochemical Screening of Ethanolic Extracts Obtained from Selected Sudanese Medicinal Plants. Current Research Journal of Biological Science. 2(2):143-146.
- Horvath, G., Kocsis, B., Botz, L., Nemeth, J., and Szabo, L. (2002). Antibacterial activity of thymus phenols by direct bioautography. Acta Biologica Szegediensis. 46: 145-6.
- Jantan, I., Yassin, M., Chin, C., Chen, L. and Sim, N. (2003). Antifungal activity of the essential oils of nine zingiberaceae species. Pharmaceutical Biology. 41: 392-397.
- Kansal, L., Sharma, A. and Lodi, S. (2011). Potential health benefits of Coriander (*Coriandrum sativum*): an overview. International Journal of Pharmaceutical Research and Development. 4(02): 10-20.
- Khan, M., Kihara, M. and Omoloso, A. (2003). Antimicrobial activity of the alkaloidal constituents of the rootbark of *Eupomatia laurina*. Pharmaceutical Biology. 41: 277-80.
- Kubo, I., Fujita, K., Kubo, A., Nihei, K. and Ogura, T. (2004). Antibacterial activity of Coriander volatile compounds against *Salmonella choleraesuis*. Journal of Agricultural and Food Chemistry. 52: 3329-3332.
- Matasyoh, J.C., Maiyo, Z.C., Ngure, R.M. and Chepkorir, R. (2009). Chemical composition and antimicrobial activity

of essential oil of *Coriandrum sativum*. Food Chemistry. 113: 526-529.

- Mildner-Szkudlarz, S., Zawirska-Wojtasiak, R., Obuchowski, W. and Gośliński, M. (2009). Evaluation of antioxidant activity of green tea extract and its effect on the biscuits lipid fraction oxidative stability. Journal of Food Science. 74: 362-370.
- NCCLS (2000). Performance standards for antimicrobial disk susceptibility tests: Approval standard M2-A7, 7th edition. Pennsylvania: Clinical and Laboratory Standards Institute.
- Perez, R.M. (2003). Antiviral Activity of Compounds Isolated from Plants. Pharmaceutical Biology. 41: 107-57
- Reynolds, J.E.F (1996). Martindale-the Extra pharmacopoeia 31st edn London Royal Pharmaceutical society of Great Britain.
- Samant, S.S., Dhar, U. and Palni, L.M.S. (1998). Medicinal Plants of Indian Himalayas: Diversity, Distribution, Potential values. Himavikas Publication No. 13. G.B. Pant Institute of Himalayan Environment and Development, Almora. Uttaranchal, India.
- Singh, G., Marimuthu, P., Murali, H.S. and Bawa, A.S. (2005). Antioxidant and antibacterial potential of essential oils and extracts isolated from various spice materials. Journal of Food Safety. 25: 130-145.
- Sreelatha, S., Padma, P.P. and Umadevi, M. (2009). Protective effects of *Coriandrum sativum* extracts on carbon tetrachloride-induced hepatotoxicity in rats. Food and Chemical Toxicology. 47: 702-708.
- Teye, G.A., Mustapha, F.B., Abu, A. and Teye, M. (2013). Effect of Moringa (*Moringa oleifera*) leaf powder and sweet basil (*Ocimum basilicum*) leaf paste on sensory and nutritional qualities of beef and ham burgers– A

preliminary study. Scientific Journal of Animal Science. 2(2): 41-46.

- Verma, A., Dhanik, J., Agarwal, D., Arya, N. and VivekaNand. (2018). Qualitative phytochemical and cluster analysis of genotypic extracts of Coriander leaves and seeds from Tarai and Kumaun regions of Uttarakhand, Himalayan state of India. International Journal of Chemical Studies. 6(2): 1566-1571.
- Verma, A., Sharma, P. and VivekaNand. (2015). Evaluation of Coriander Extracts of Leaves and Seeds Genotypes against the Feeding Activity of 8 Day Old Larvae *Spodoptera Litura* (Fab.). Asian Resonance. 4 (3): 69-74.
- Zoubiri, S. and Baaliouamer, A. (2010). Essential oil composition of *Coriandrum sativum* seed cultivated in Algeria as food grains protectant. Food Chemistry. 122: 1226-1228.

How to cite this article:

Ankita Verma, Deepali Agarwal, Jyotsna Dhanik, Neelam Arya and Viveka Nand. 2018. Antibacterial Efficacy and Cluster Analysis of Genotypic Extracts of Coriander Leaves and Seeds against Human Pathogenic Bacterial Strains. *Int.J.Curr.Microbiol.App.Sci.* 7(05): 2727-2736. doi: <u>https://doi.org/10.20546/ijcmas.2018.705.316</u>